

## Fermentation technologies for the production of exopolysaccharide-synthesizing *Lactobacillus rhamnosus* concentrated cultures

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**Financial support:** This work was carried out within the Research Network on Lactic Acid Bacteria, supported by the Natural Sciences and Engineering Research Council of Canada, Agriculture and Agri-food Canada, Novalait Inc., The Dairy Farmers of Canada and Rosell-Lallemand Inc.

**Keywords:** alginate beads, fed-batch culture, temperature.

**Abbreviations:** AS: automated spectrophotometry  
CFU: colony forming units  
CRV: capillary relative viscosities  
EPS: exopolysaccharide  
ICT: immobilized cell technology  
LAB: lactic acid bacteria  
MMRS: modified-MRS  
OD: optical density  
SEM: Standard error of the means  
WP: whey permeate

The exopolysaccharide (EPS)-producing cultures such as *Lactobacillus rhamnosus* RW-9595M present a challenge for the culture producers because the high viscosity of the fermented growth medium makes it difficult to recover the cells by centrifugation or filtration. This study examined four approaches to reduce viscosity of the medium while producing high cell densities: incubation temperature, extended incubation in the stationary growth phase, production in alginate gel beads and fed-batch fermentation technology. Automated spectrophotometry (AS) was used to study the effects of temperature, pH and lactate level on growth of the strain. In AS assays, there was no significant difference in final maximal biomass

production at temperatures ranging between 34°C to 44°C, but lower yields were noted at 46°C. A pH below 6.0 and a lactate concentration higher than 4% almost completely prevented growth. Under batch fermentation conditions, the viscosity of the medium obtained at 37°C was two fold higher than for 44°C. For cultures produced at 37°C, centrifugation at 10000 g during 5 min did not allow complete recovery of cells, in contrast to cultures grown at 44°C. An extended period of incubation (5 hrs) in the stationary growth phase did not reduce the final viscosity of the growth medium. For similar biomass levels, the glucose-based fed-batch fermentation allowed a 40% reduction in viscosity of the fermented medium in comparison to traditional batch

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**cultures. High-density cell populations ( $3 \times 10^{10}$  CFU/g) were obtained when *L. rhamnosus* RW-9595M was grown in alginate beads. However, overall biomass yields in the immobilized cell bioreactor were half of those obtained in free-cell fermentations. Therefore three methods of producing concentrated EPS-producing cultures are proposed.**

Exopolysaccharide (EPS)-producing lactic acid bacteria (LAB) are of interest to the food industry because they can improve the texture of yoghurts (De Vuyst and Degeest, 1999; De Vuyst et al. 2001), decrease the risk of syneresis and improve yields in cheeses (Broadbent et al. 2001). In addition to their technological benefits, LAB EPS have been recognized to have antitumoral, antiulcer and blood cholesterol lowering activities, as well as the ability to enhance the immune system (Chabot et al. 2001; Ruas-Madiedo et al. 2002). Therefore EPS-producing strains are of commercial value for both their technological and putative probiotic properties.

In order to supply cultures directly to consumers or to the food industry, manufacturers must propagate the strains in appropriate media, recover the cells and stabilize cell concentrates by freezing or drying. With high EPS producing bacteria, such as *Lactobacillus rhamnosus* RW-9595M (Macedo et al. 2002; Bergmaier et al. 2003), cell recovery presents a challenge for manufacturers because viscosity of the fermented medium could hinder centrifugation or filtration operations. There is therefore a need to develop fermentation technologies which can generate high biomass yields, while keeping the viscosity at a level which would enable easy recovery of the cells from the fermented broth.

A balance between carbon and nitrogen sources is necessary to obtain a high EPS concentration (De Vuyst and Degeest, 1999), but other factors such as growth temperature, pH and agitation affect EPS production (Gamar-Nourani et al. 1998). High incubation temperatures have been shown to reduce EPS production by lactobacilli (Grobben et al. 1995; Mozzi et al. 1996b), but it is unknown if the viscosity reduction is sufficient to enable easy recovery of cells. Fed-batch fermentation with high carbohydrate levels was successful in enhancing EPS production (Cheirsilp et al. 2003) but is it unknown if a reverse strategy (*i.e.* fed-batch with low carbohydrate level) could be used to generate a fermented medium with high cell concentration and low-viscosity.

In addition to the traditional concentrated starter technology, immobilized cell technology (ICT) has been proposed for the production of bacterial concentrates (Doleyres and Lacroix, 2005). Although the production of concentrates in alginate beads has the advantage of eliminating the need for a concentration step, the EPS yields are sometimes lower (Champagne et al. 1993) and some strains are adversely affected by entrapment in alginate gels (Lamboley et al. 2003). No data are available

on the ICT production of EPS-producing strains of *L. rhamnosus*.

The objective of the present study was to examine four methods for the production of high bacterial populations while controlling viscosity of the medium: high incubation temperatures, extended incubation on the stationary growth phase (SGF), fed-batch fermentations and biomass production in alginate beads.

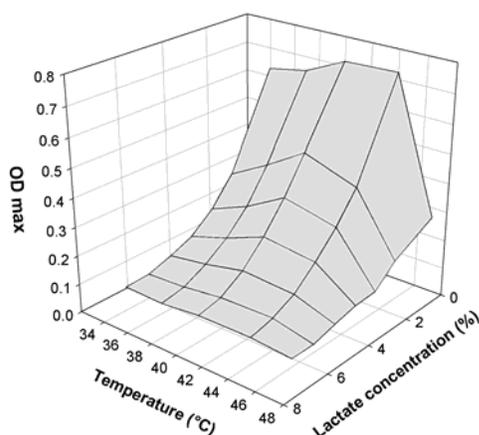
## MATERIALS AND METHODS

### Cultures

Stock cultures of *L. rhamnosus* RW-9595M were prepared by mixing a fresh MRS-grown culture with sterile rehydrated skim milk (20% w/w) and sterile glycerol (20% w/w) in a 1:2:2 ratio, and placing 1 ml fraction in Cryovials (Nalge, Rochester, USA). The cell suspensions were then frozen and kept at  $-70^{\circ}\text{C}$  until used. The inocula for fermentations were prepared by adding 1 ml of a thawed stock culture to 50 ml MRS broth and incubating at  $37^{\circ}\text{C}$  until a pH of 4.5-4.8 was reached (typically after 6.5 hrs). For spectrophotometric assays, 1 ml of MRS-grown culture was added to 9 ml of 0.01% (w/v) sterile peptone water (Difco, Detroit, USA) and this cell suspension served to inoculate the wells in the microplates. For the preparation of the alginate-entrapped culture, a fresh MRS-grown culture (15 ml) was mixed with 60 ml of a sterile 2% (w/v) alginate solution (Sobalg 126, Grindsted, Denmark), and the cell suspension was added dropwise to 200 ml of a 0.2 M  $\text{CaCl}_2$  solution. Beads with approximately 2 mm diameter were left in the  $\text{CaCl}_2$  solution for 30 min to allow hardening and then recovered in a sterile Buchner filter. Approximately 62 g of beads were obtained.

### Spectrophotometric analyses for the determination of growth parameters

Since Method 1 was based on the use of over-optimal growth temperatures, it was necessary to ascertain the optimum growth temperature for this particular strain. With respect to Method 4, the strategy required prior knowledge of the amount of sugar needed to carry out an extended fermentation. Thus, preliminary assays using automated spectrophotometry (AS) were carried out to ascertain the conditions of pH, temperature and maximum lactate concentration that were to be subsequently used for fermentations. The AS assays were not conducted on the modified-MRS (MMRS) or whey media used for fermentations because such media generate biomass levels which are too high. Indeed, in AS assays it is recommended to limit the growth of the culture to optical density (OD) values under 1.0. Thus AS assays were conducted in the basal medium described by Morishita et al. (1981) for lactobacilli. It had the following composition per liter: 70 g lactose, 3 g  $\text{KH}_2\text{PO}_4$ , 3 g  $\text{K}_2\text{HPO}_4$ , 1 g Tween 80 and 1 ml of a concentrated stock solution of the following elements to a final concentration per litre of: 0.05 g  $\text{MnSO}_4 \cdot 7\text{H}_2\text{O}$ ,



**Figure 1. Effect of growth temperature and initial lactate concentration on maximal optical density (OD max) of *Lactobacillus rhamnosus* RW-9595 M in the medium used in AS assays.**

0.5 g  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 6 g sodium acetate, 1 g ammonium citrate, 0.02 g  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ . Then, 100 ml of a stock solution of amino acids composed per liter of :1 g L-arginine, 2 g L-aspartic acid, 2 g L-cysteine, 2 g L-glutamic acid, 1 g L-leucine, 1 g L-isoleucine, 1 g L-lysine, 1 g L-methionine, 1 g L-phenylalanine, 1 g L-serine, 1 g L-threonine, 1 g L-tryptophan, 1 g L-tyrosine, 1 g L-valine, were added. For the pH assays, the pH was adjusted to 5.1, 5.4, 5.7, 6.0 and 6.3 with HCl 3M prior to sterilisation at 121°C for 5 min. This was the actual holding time of the medium at 121°C. The holding time was initiated when the medium had reached 121°C and not when the sterilization chamber had attained the temperature, which is much earlier. This was enabled by placing a temperature probe in the medium during sterilization. After cooling, 1 ml per liter of a filter sterilized stock solution of vitamins (per litre composed of 0.1 g folic acid, 1 g nicotinic acid, 1 g pantothenic acid, 2 g pyridoxal and 1 g riboflavine) completed the medium formulation. For the lactate concentration assays, a solution of 60% sodium lactate (Fisher, Montréal, QC, Canada) was added to the base medium to obtain a final lactate concentration of 0, 2, 3, 4, 5, 6 and 7% (w/v). The pH was adjusted to 6.0 prior to sterilisation.

200  $\mu\text{l}$  of sterile media were then dispensed in wells (Polystyrene sterile 96 well plates, Costar, NY, USA) and inoculated with 10  $\mu\text{l}$  of fresh inoculum. Noninoculated wells were also used to determine the baseline of each condition. The plates were then placed into a Powerwave (KC-4, Bio-Tek, VT, USA) incubation chamber for which shaking frequency (10 sec every 15 min), intensity (low), wavelength (600 nm) and OD reading frequency (every 15 min) were kept constant for the following temperatures: 34,

37, 40, 44 or 46°C. In each assay, triplicate wells for each condition were performed and mean values were used to calculate the maximal optical density (OD max) and the maximal growth rate ( $\mu$  max) for the given assay. Three independent replications were carried out.

### Method 1. Use of over-optimal growth temperatures

In assays with Method 1, the growth medium was selected to generate the highest possible biomass and, hence, a high EPS level. This was done to simulate industrial conditions which are designed to attain high biomass, as well as to generate the highest possible viscosity in order to test the value of our hypothesis. The medium used to evaluate the effect of temperature on cell growth and viscosity was formulated with ingredients of commercial sources recognized as enabling very good growth of strain RW-9595M as well as high EPS levels (Macedo et al. 2002). The whey permeate (WP) based medium was prepared by first adding 78 g WP powder (Saputo, Montréal) in 800 ml of distilled water. The pH was adjusted to 5.0 with HCl 3M prior to heating (121°C, 10 min). After cooling, the solution was filtered on a 8  $\mu\text{m}$  Whatman (MI, USA) #2 paper to remove heat-formed precipitates. The pH was then readjusted to 5.0 and the following solution added per litre: 2.5 ml Tween 80, 3 g  $\text{KH}_2\text{PO}_4$ , 3 g  $\text{K}_2\text{HPO}_4$ , 100 ml of the stock solution of mineral salts, 100 ml of amino acids as described above and 5 g of yeast extracts previously ultrafiltered at 10 kDa to remove polysaccharides (Champagne et al. 1999). The medium was sterilised again (121°C, 1 min), cooled to room temperature and 1 ml of the stock solution of vitamins was added per litre of medium.

Batch cultures were conducted in three 2.5 L fermentors (New Brunswick Scientific Co., models BioFlo III and BioFlo 3000, Edison, NJ, USA) inoculated at 0.6% (v/v) with a fresh sub-culture at a controlled temperature of 37, 42 or 44°C, with agitation set at 100 rpm and pH controlled at 6.0 by addition of a mixture of KOH 6N and  $\text{NH}_4\text{OH}$  6N prepared in 5:1 ratio. Fermentations were carried out until stabilisation of the OD. Samples were aseptically withdrawn when the SGP had been reached (between 11.5 and 14.2 hrs; Table 1) to determine bacterial populations, lactose consumption and lactic acid production, as well as relative viscosity of the culture broth.

### Method 2. Over-incubation of the culture in the SGP

In Method 1, samples were taken at the beginning of the SGP. For Method 2, the incubation of the same media was simply extended at 42 or 44°C for 5 hrs in the SGP. Samples were then taken again to ascertain the effect of this extended incubation on viable counts, viscosity and OD before and after centrifugation. Sampling times varied between 16.5 and 17.8 hrs (Table 1) as a function of the time at which the SGP had been reached.

**Table 1. Effect of fermentation temperature and extended incubation in the stationary growth phase (5 hrs) on biomass levels of *Lactobacillus rhamnosus* RW-9595M, medium CRV and the subsequent removal of cells by centrifugation.**

Fermentation		Fermentation time (hr)	Population (CFU/ml)	Cells per chain <sup>1</sup>	CRV	OD		
Temperature	Growth phase					Before CF <sup>2</sup>	After CF 4000 g	After CF 10000 g
37°C	Beginning stationary	14.2 (0.4)	9.3 (0.9) x 10 <sup>9</sup>	1.3 (0.1)	4.81 (0.29)	11.1 (0.1)	5.7 (0.1)	5.8 (0.1)
42°C	Beginning stationary	12.8 (0.3)	7.2 (1.3) x 10 <sup>9</sup>	1.7 (0.1)	2.66 (0.27)	11.8 (0.3)	5.9 (0.1)	1.7 (1.1)
42°C	5 hrs in stationary	17.8 (0.3)	5.7 (0.6) x 10 <sup>9</sup>	1.8 (0.2)	2.97 (0.32)	11.4 (0.3)	5.8 (0.1)	2.4 (1.0)
44°C	Beginning stationary	11.5 (0.6)	6.3 (1.1) x 10 <sup>9</sup>	1.5 (0.3)	2.20 (0.06)	12.0 (0.2)	5.8 (0.1)	0.1 (0.1)
44°C	5 hrs in stationary	16.5 (0.6)	5.4 (0.6) x 10 <sup>9</sup>	2.0 (0.4)	2.25 (0.04)	11.7 (0.3)	5.7 (0.1)	0.1 (0.1)

<sup>1</sup> Average number of cells per chain

<sup>2</sup> CF: Centrifugation

Values in brackets represent SEM obtained from three independent assays.

### Method 3. Production in alginate beads

Assays with Method 3 and Method 4 were done simultaneously in order to compare biomass levels of the three fermentation technologies (free, immobilized and fed-batch). However, the WP medium could not be used for Method 4, because it was necessary to have a low level of carbohydrate at the beginning of the fermentation (low C/N ratio), which the WP media did not enable. It was therefore necessary to use a synthetic medium where the carbohydrate could be added separately from the other ingredients. A MMRS was thus formulated for its carbohydrate-defined composition as opposed to the WP-based medium where the lactose content cannot be varied without changing the other solids content. The MMRS was prepared as follows (per litre of medium): 10 g tryptone (N1, Organotechnie, TechniScience Montreal QC Canada), 10 g beef extract, 5 g yeast extract (FNI 100, Lallemand, Montreal QC Canada), 1 g Tween 80, 1 g ammonium citrate, 2 g sodium acetate, 0.1 MgSO<sub>4</sub>, 0.05 g MnSO<sub>4</sub>, 1 g K<sub>2</sub>HPO<sub>4</sub>. The medium (1.5 litre) was sterilized at 121°C for 10 min (actual temperature probe-controlled holding time at 121°C). Solutions with 60% glucose (w/v) or 50% CaCl<sub>2</sub> (w/v) were prepared separately, and sterilized at 121°C for 10 min. In the control (batch) fermentation with free cells, the fermenter jar was sterilized at 121°C for 30 min and sterile MMRS was added. The MMRS base (1.5 litre) was supplemented with 180 ml of the 60% glucose solution and 50 ml of 50% CaCl<sub>2</sub>, for final concentrations of 0.1 M of CaCl<sub>2</sub> and 60 g/l glucose. This medium, with all the glucose

added at the beginning of the fermentation, was used for the free cell control as well as for the alginate bead-based ICT method. The medium was inoculated with 15 ml of an MRS-grown culture mixed with 47 ml of 0.2 M CaCl<sub>2</sub>.

This inoculation procedure was carried out in order to compare this treatment with batch fermentations performed with alginate beads, where the same volume of fresh culture was entrapped in 62 g of beads recovered from the 0.2 M CaCl<sub>2</sub> solution. Bead samples were taken for analyses at the beginning of the SGP (when acidification has stopped).

With alginate beads, fermentations inoculated with 62 g of the freshly prepared alginate particles were carried out. The medium was supplemented with 50 mL of 50% CaCl<sub>2</sub> to maintain bead integrity. The fermentation and samplings were carried out as for the free cell batch fermentations. Viable populations in the beads as well as in the medium, capillary relative viscosities (CRV) of the medium and sugar/acid concentrations were analysed.

### Method 4. Use of fed-batch technology

In fed-batch fermentations, the MMRS base (1.5 litre) was supplemented with 50 ml of the concentrated (50%) CaCl<sub>2</sub> solution and only 30 ml of 60% glucose solution for a starting concentration of 1.2%. Inoculation and fermentation were conducted as described for the free-cell control until acid production stopped, indicating complete assimilation of the initial glucose. Injection of sterile 60% glucose at a 30 ml/h rate was then carried out over 5 hrs.

Samples were taken when the fed batch process was initiated, as well as at every hr thereafter. Viable populations, CRV of the medium and sugar/acid concentrations were analysed.

All fermentations (batch or fed batch) were carried out on 2.5 litre New-Brunswick Scientific BioFlo 3000 or BioFlo III units. The temperature was maintained at 42°C, agitation set at 100 rpm, and pH kept at 6.0 by addition of a mixture of KOH 6N and NH<sub>4</sub>OH 6N prepared in 5:1 ratio.

### Microbial analyses

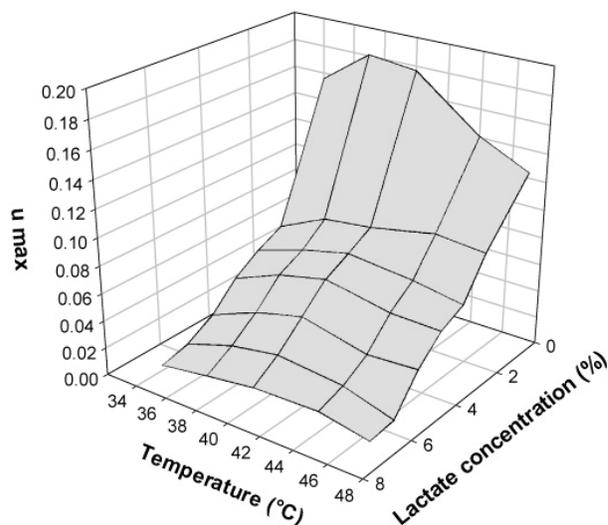
The viable population in the medium (free cells) was determined by plating appropriate dilutions of the cultures on MRS agar, and incubating at 37°C for 48 hrs. To break cell chains, the cell suspension of the first dilution (1 ml of culture in 99 ml of 0.1% peptone) was treated with a high-speed (25000 rpm) homogenization probe (Omni International, Marietta GA USA) during 1 min. The viable population in the beads was determined by dissolving approximately 0.5 g of beads in 99 ml of 1% sodium citrate, and plating the subsequent dilutions in 0.1% peptone as described above. The first dilution bottle was then also treated by the high-speed homogenization to break cell chains. Since lactobacilli grow in cell chains of various lengths, colony forming units (CFU) counts may underestimate of the actual population values. Indeed, if chains have, on the average, 3 cells per chain, then the CFU counts only register 33% of the actual cell population. Therefore, in order to try to accurately present the true number of cells in the sample, we examined culture smears at a 1000 X enlargement ocular and tried to assess the average number of cells per chain. To provide total cell counts, we then simply multiply this “average number of cells per chain” with the CFU count. The cells were coloured with methylene blue prior to examination.

### HPLC analyses

Substrates and products were analysed by HPLC using a Waters (Mississauga, Canada) system coupled to Millennium software. Samples were centrifuged and filtered on a HVLP 0.22 : membrane prior separation on a Aminex HPX 87H column (BioRad, Mississauga, Canada) heated at 45°C. Detection was done with a refractive index monitor (model 410, Waters) controlled at 40°C and a photodiode array detector model 996 (Waters). The mobile phase was made of H<sub>2</sub>SO<sub>4</sub> 0.008N and circulated at 0.4 ml/min.

### OD measurements in the fermented media

Growth media were centrifuged at 4000 g or 10000 g during 5 min at 4°C and samples analysed for OD before and after (supernatants) centrifugation to estimate the remaining biomass after centrifugation. Since the relationship between biomass and OD is not linear with OD values above 1.0, samples having high OD needed to be



**Figure 2. Effect of growth temperature and initial lactate concentration on maximal growth rate ( $\mu$  max) of *Lactobacillus rhamnosus* RW-9595M.**

diluted prior to reading. The OD readings were carried out by placing the sample (diluted or not) in a 1 ml cuvette, which was scanned using a Beckman, DU7400 (CA, USA) spectrophotometer. The final OD was obtained by multiplying the diluted OD by the dilution factor when necessary.

### Capillary relative viscosity determination

15 ml of broth of culture or supernatant were introduced in a capillary Ubbelohde 1B unit placed into a water bath with a control temperature at 25°C for 10 min. With the capillary unit used in these assay, 15 ml of water typically required 19 sec to flow out of the system. The CRV reported are the ratio between the time (in sec) required by the experimental sample to flow out of the viscometer and that of water at the same experimental conditions. Reported data are the mean of triplicate analysis.

### Statistical analyses

Data are the average of three independent assays. CFU values were converted into their corresponding Log<sub>10</sub> numbers and ANOVA were then carried out using the Student-Newman-Keuls procedure with InStat 3.0 software (GraphPad, San Diego, USA). The  $\alpha = 0.05$  level was used to ascertain the statistical significance of the differences. Paired t tests were used to evaluate the effect of extended incubations on viability drops at each incubation temperature (Method 2). Standard Error of the Means (SEM) values are presented in brackets in the tables.

## RESULTS AND DISCUSSION

Many studies show that EPS production in lactobacilli is strongly linked to biomass levels (De Vuyst et al. 1998;

Torino et al. 2000). Although viscosity does not always correlate with EPS production (Shihata and Shah, 2002; Ruas-Madiedo et al. 2005) for *L. rhamnosus* RW-9595M, there is a correlation between EPS concentration in the medium and CRV values (Macedo et al. 2002). Then, by keeping the biomass yield of a culture at a low level, one could achieve the goal of having a low-viscosity fermented medium. However, from a starter manufacturer point of view, high biomass productions are desirable, which eventually lead to high medium viscosity. Attempts were thus made to find fermentation parameters which would enable high biomass as well as low EPS production.

### **Selection of temperature, lactate concentration and pH parameters for fermentation assays**

AS evaluates growth from continuous OD readings, and media must be specially formulated for these assays in order to limit the growth in an OD range in which OD and biomass are linearly correlated. AS enabled the evaluation of the combined effects of lactate concentration, pH and temperature on the growth of *L. rhamnosus* RW-9595M.

ANOVA on OD max data from media at pH 6.0 with no added lactate (0% lactate, Figure 1) revealed that there was no significant ( $P > 0.05$ ) effect of incubation temperatures between 34 and 44°C on OD max values, but that 46°C was detrimental to biomass production.

Data for  $\mu$  max differed from that of OD max. At pH 6.0 and without added lactate (0% lactate, Figure 2), the ANOVA revealed that the highest growth rates ( $\mu$  max values) were obtained between 34 and 40°C and decreased significantly at higher temperatures (Figure 2). These data enabled the selection of 37°C, 42°C and 44°C for fermentation assays on the effect of temperature (Method 1). Incubation at 46°C clearly affected both biomass yield and growth rate and was unacceptable. Although the growth rate at 44°C was lower than at 40 or 37°C (Figure 2) the biomass levels were similar for the three temperatures. Thus, 44°C was selected as the over-optimal growth temperature for the assays in Method 1.

When pH of the media were adjusted below 6.0, the ANOVA analysis of  $\mu$  max data in media with no added lactate revealed significantly lower values (data not shown). These data served to select a pH value of 6.0 for subsequent biomass productions in fermenters. These results are in line with those of Mozzi et al. (1996a) who reported lower populations under pH 6.

Lactate concentrations higher than 4% substantially decreased the growth of *L. rhamnosus* RW-9595M, and negligible growth was noted in media adjusted at pH 6.0 having 6% lactate (Figure 1 and Figure 2). These AS results thus enabled the selection of 6% of glucose for the fed-batch assays (Method 4), in order to generate the 4% lactate level judged to be critically detrimental to biomass and growth rate levels. Indeed, preliminary data with this strain

(not shown) suggested a conversion level of glucose to lactate of approximately 75% (Table 2).

Under pH control, lactate accumulation can generate growth inhibition (Cachon and Diviès, 1993), but the inhibitory level varies between strains, species, pH and temperature (Houtsma et al. 1996). The AS data was in line with the literature, showing lactobacilli to be more tolerant to lactate than are lactococci (Stieber et al. 1977; Cachon and Diviès, 1993).

### **Method 1. Effect of temperature on biomass and viscosity of the fermented media**

During pH-controlled batch productions in the whey-based medium, fermentation times to reach the SGP were longer for temperatures below 42°C (Table 1), which was in line with the AS data (Figure 1 and Figure 2). The highest CFU/ml value was obtained at 37°C (Table 1) but the ANOVA on data at the beginning of the SGP showed that the difference with those at 42 and 44°C was not judged to be significant ( $P > 0.05$ ) when the average number of cells per chain value was taken into account and CFU values were multiplied by these values (Table 1) to ascertain the total cell counts. The populations obtained were in accordance with those reported by Bergmaier et al. (2003) on WP medium after 18 hrs of fermentation for this *L. rhamnosus* RW-9595M strain.

CRV of the media was 2-fold higher at 37°C than at 42°C or 44°C (Table 1). These data are in agreement with many studies showing higher viscosities at sub-optimal growth temperatures (Gassem et al. 1995; Mozzi et al. 1995) as well as with those of Gamar-Nourani et al. (1997) who found lower EPS production by *L. rhamnosus* at high incubation temperatures. Under these conditions, because of the higher viscosity obtained at 37°C, even a centrifugation at 10000 g for 5 min did not allow complete retrieval of the biomass from the growth medium (Table 1).

Complete retrieval of cells was not possible for any of the cultures when using a centrifugation force of 4000 g during 5 min. However, at 10000 g, the low viscosity of the culture grown at 44°C allowed complete separation of the bacteria from the medium. Therefore, this study was successful in showing that over-optimal incubation temperatures can be used to lower medium viscosity sufficiently to recover cells under certain centrifugation conditions. It remains to be determined to what extent this strategy can be used for other species. Indeed, with many thermophilic strains, EPS production (and hence viscosity) seems to be growth-associated (De Vuyst et al. 1998).

### **Method 2. Extended incubation**

Since it was reported that EPS production (Pham et al. 2000) and CRV (Macedo et al. 2002) of *L. rhamnosus* cultures were at its highest at the beginning of the SGP and dropped steadily thereafter, an attempt was made to see if

**Table 2. Effect of fermentation technologies on the population of *Lactobacillus rhamnosus* RW-9595M, medium CRV and glucose/lactate levels in the fermented media.**

Fermentation technology	Population (CFU/ml or /g)	CRV	Glucose (g/l)	Lactate (g/l)
<b>Free cell control</b>	3.0 (0.9) x 10 <sup>9</sup>	5.07 (0.32)	0	37.3 (3.2)
<b>Immobilized cells</b>				
Beads	3.0 (1.5) x 10 <sup>10</sup>	4.41 (0.92)	0.4 (0.03)	38.7 (2.6)
Free cells outside beads	1.1 (2.4) x 10 <sup>9</sup>			
<b>Fed batch</b>				
Beginning	8.3 (2.0) x 10 <sup>8</sup>	1.26 (0.17)	0	9.7 (1.7)
1 hr	8.0 (1.9) x 10 <sup>8</sup>	1.52 (0.18)	0.9 (0.1)	16.0 (1.8)
2 hrs	2.0 (0.9) x 10 <sup>9</sup>	1.60 (0.30)	2.5 (0.6)	21.6 (3.0)
3 hrs	2.1 (0.2) x 10 <sup>9</sup>	2.23 (0.13)	2.5 (0.3)	22.5 (1.3)
4 hrs	2.6 (0.4) x 10 <sup>9</sup>	2.50 (0.42)	5.6 (0.4)	33.4 (4.2)
5 hrs	3.9 (1.4) x 10 <sup>9</sup>	3.25 (0.29)	7.9 (0.5)	38.7 (2.9)

Values in brackets represent SEM obtained from three independent assays.

this phenomenon could be used to enhance recovery of the cells by centrifugation. Unfortunately, the CRV of the fermented medium was not significantly reduced after 5 hrs extended incubation in the SGP (Table 1). It was suggested that hydrolases activated by specific growth conditions caused degradation of the EPS (Pham et al. 2000), but our experimental conditions do not seem to have generated this EPS hydrolysis.

An apparent drop in viable counts of approximately 20% was noted during prolonged fermentation periods, but paired t tests on population differences are not shown to be significant when total populations are calculated (multiplication of the CFU vales by the average number of cells per chain).

### Method 3. ICT

In the ICT system, populations per g of beads in the MMRS were 10 times higher than for the free-cell control batch fermentations (Table 2). However, when the total yield of the bioreactor is considered, the total population from the beads represented only 40% of that obtained in free cells bioreactor. CRV values obtained for immobilized cells were slightly lower than for the free cells. Lactate concentrations for both treatments were similar.

The production of biomass by growing lactic cultures in alginate beads had previously been suggested (Champagne et al. 1992; Champagne et al. 1993), particularly for cultures which were sensitive to oxygen or centrifugation stresses. For EPS producers, the immobilization techniques may facilitate the recovery of the cells even in a viscous environment as they are entrapped in beads (Table 2) and this presents the main advantage. Thus, recovery of the beads could easily be carried out with a simple filtration unit. However, an important disadvantage with this technology was the lower biomass yields. A method to improve populations in beads, and reduced cell release into the medium, must be developed.

At 42°C, the free cell control produced on MMRS (Table 2) yielded a lower total biomass than the one obtained from the supplemented WP (Table 1). On the other hand, the viscosities obtained with the MMRS medium with the free and immobilized cells were higher to the ones obtained in the supplemented WP at 42°C.

### Method 4. Fed-batch technology

During fed-batch cultures, the medium viscosity increased in parallel with the cell counts (Table 2). However, the final CRV of 3.25 was almost 40% lower than the viscosity obtained during control batch fermentation (Table 2) and

for equal biomass levels. At the end of the culture, glucose concentrations were low (0.4 g/l) and the lactate concentration was at 38.7 g/l for both the immobilized and free cells.

The quantity of carbohydrates in the medium affects EPS yields (Prasher et al. 1997) and high initial carbohydrate levels tend to enhance final EPS levels (De Vuyst et al. 1998; Cheirsilp et al. 2003; Korakli et al. 2003). It was therefore our strategy to add glucose at a level allowing maximum growth, and to prevent an excess of carbohydrate which would result in enhanced EPS production, particularly in the fed-batch assays. As mentioned previously, a 6% glucose level was chosen using the AS data. As expected, lower CRV were obtained for the fed batch productions, during which glucose concentration was equal or lower than 12 g/l. However similar final lactate concentrations and populations as for the batch production were obtained. The excess of glucose at the end of the fed batch process indicates that the feeding rate of glucose should be lowered towards the end of the fermentation. Because a lower viscosity was obtained in the fed-batch MMRS cultures, compared to control batch fermentations, it can be expected that cell recovery by centrifugation at 10000 g would be improved with this strategy, as shown in Table 1 for other conditions yielding similar CRV.

The current formulation of MMRS is expensive and other formulations could be used commercially. If defined media are used, these data show that it would be useful to add all the nitrogen-based compounds (peptones, yeast extracts) at the beginning of the fermentation and add the carbohydrate under fed-batch.

## CONCLUDING REMARKS

This study showed the effects of different culture conditions on growth of *L. rhamnosus* RW-9595M and on viscosity of fermented media. Lower viscosities and good growth were obtained by maintaining the growth temperature at 44°C and pH at 6.0 or by conducting the fermentation under a fed-batch process. Both methods could facilitate separation of the cells from the growth medium. Immobilization in alginate beads also enables the recovery of concentrated EPS-producing cultures, but the yields are lower than with free cells.

This study shows that many approaches are possible for the production of concentrated suspensions of valuable probiotic or starter EPS-producing cultures, which can be adapted to the equipment and media available or the strains involved.

## ACKNOWLEDGMENTS

Gratitude is expressed to Ms. Cathy St-Laurent for technical assistance.

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**Champagne, C.P. et al.**

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